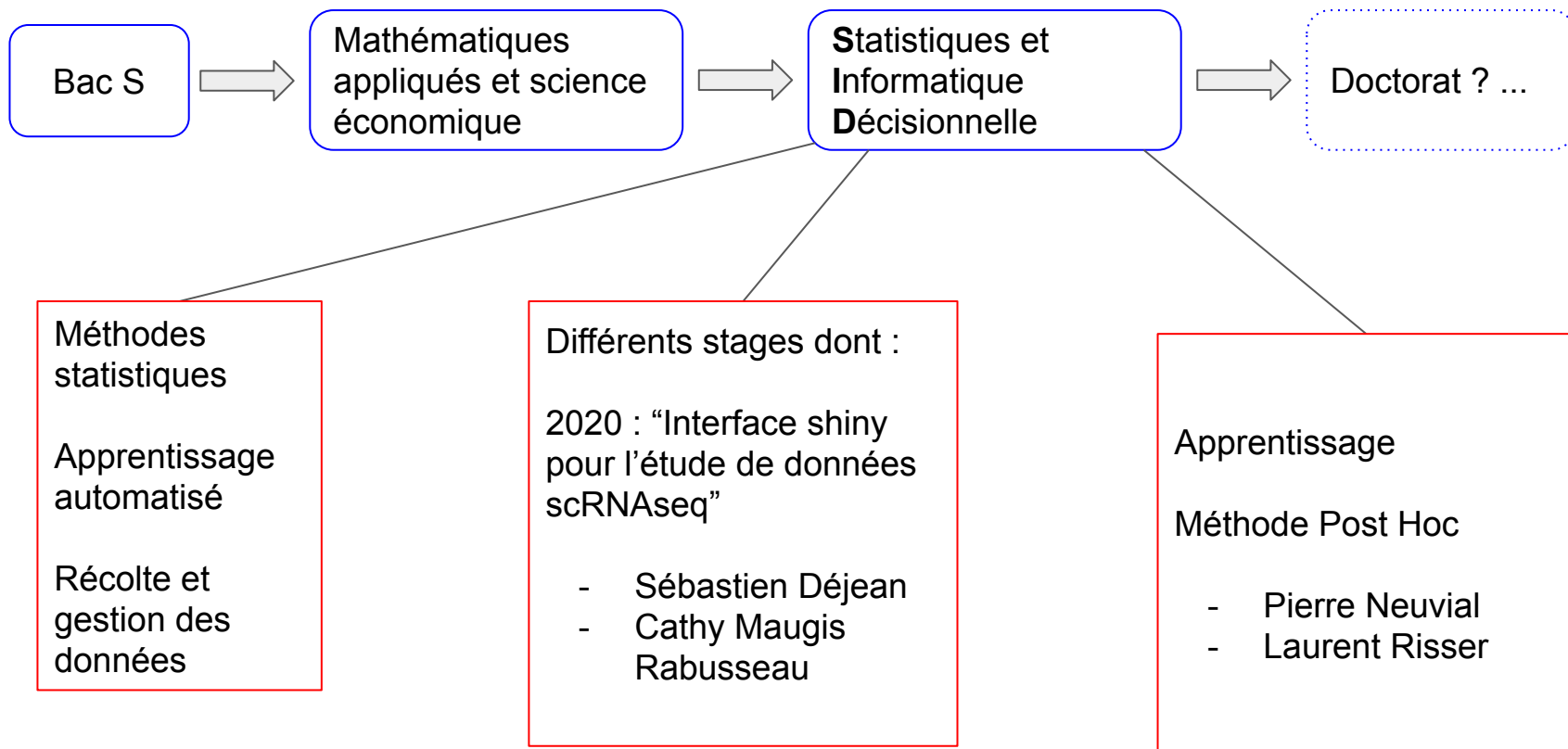


# A Shiny application To Explore Clusterings of Single-Cell RNA-seq data

ASTEC-sc

# Parcours



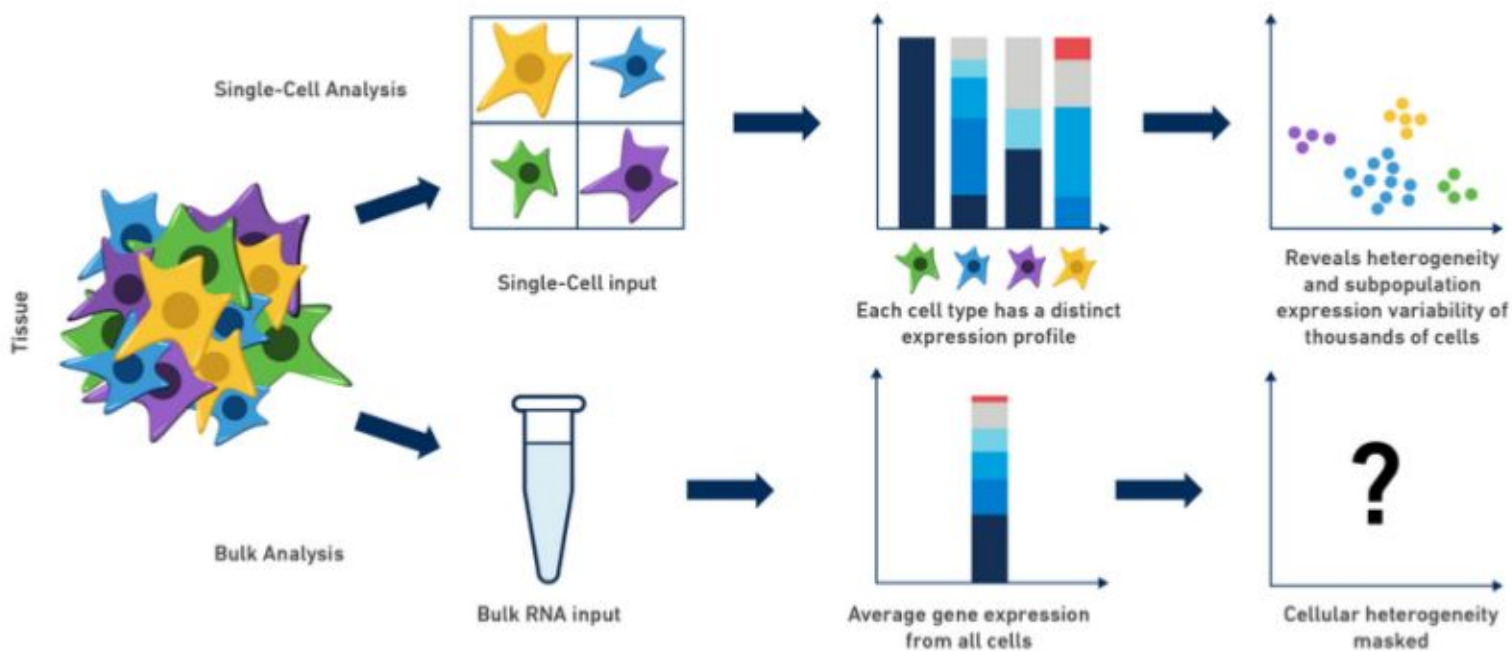
# Contents

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2. Overview of the app ASTEC-sc
3. What's next ?

# Contents

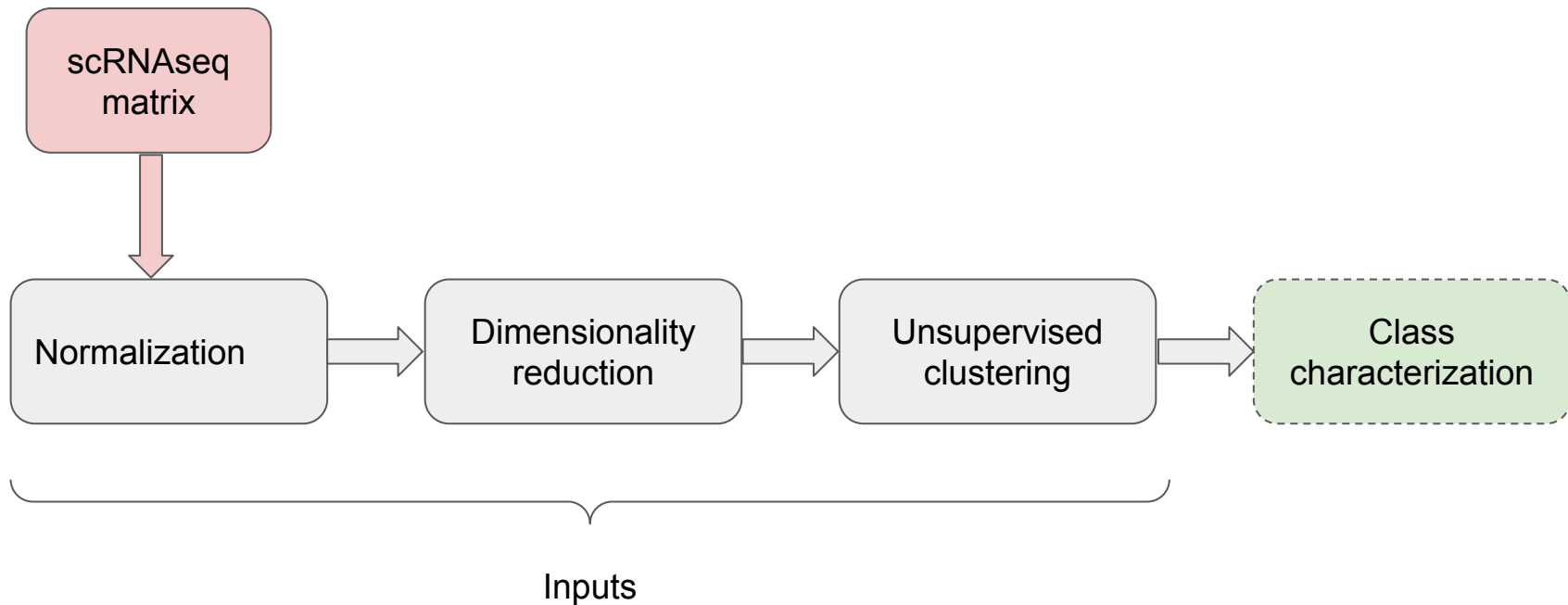
1. Context
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# Data : single cell RNA seq



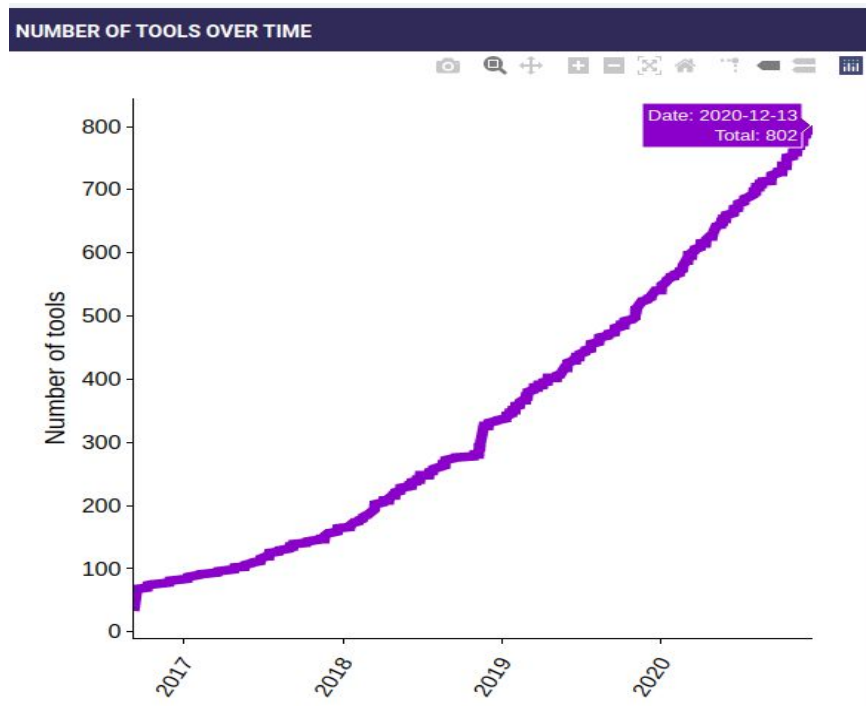
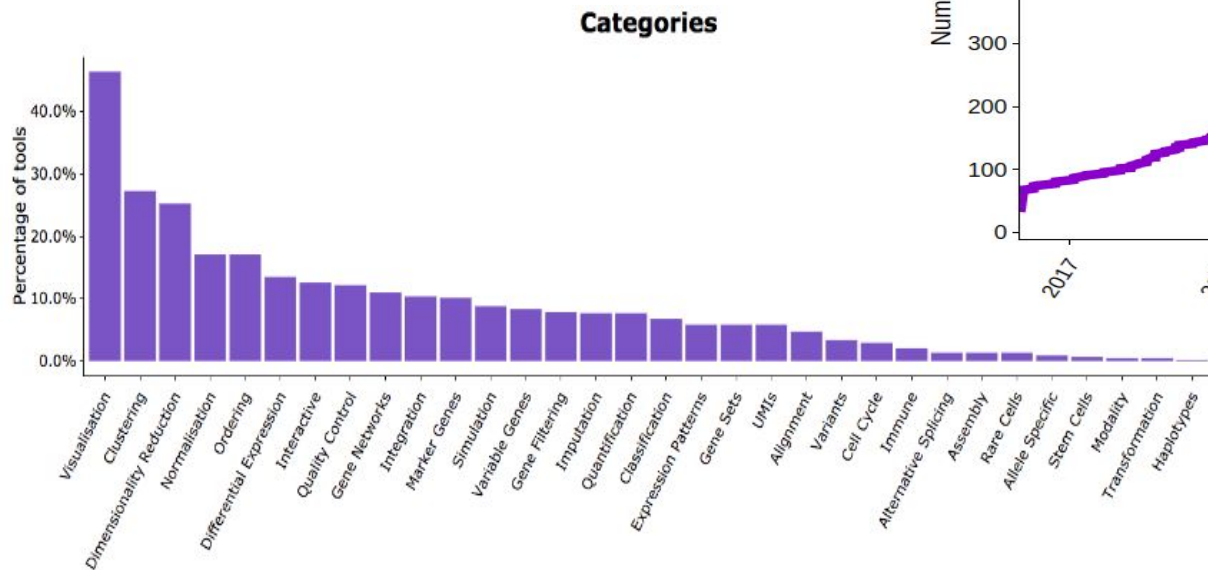
(<https://community.10xgenomics.com>)

# Main steps in statistical analysis

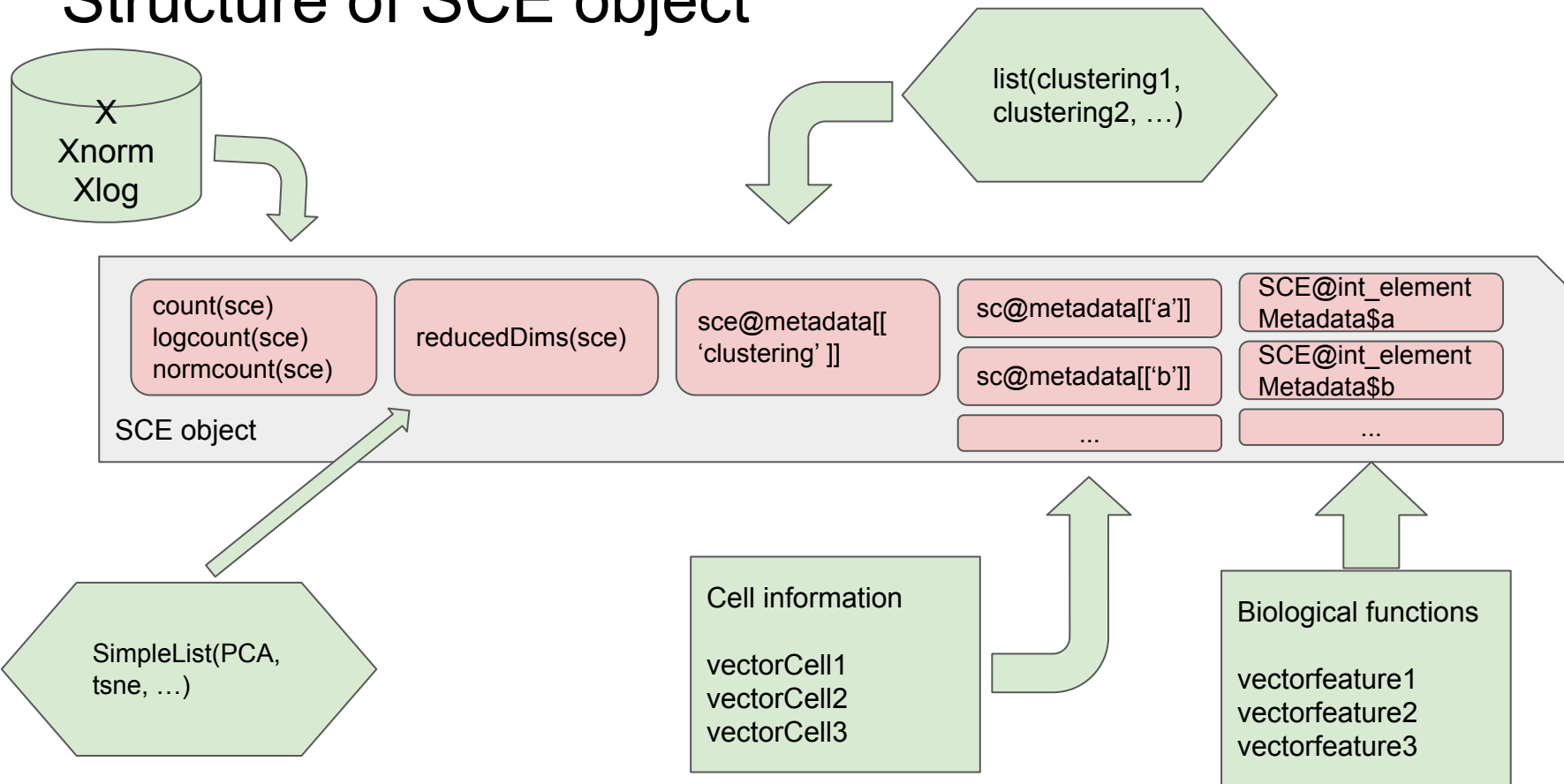


# Single cell RNA seq in R

Packages : *SingleCellExperiment (SCE)*,  
Seurat, ...



# Structure of SCE object





# Structure of SCE object

```
SCE = SingleCellExperiment( assays = SimpleList(counts=X, normcounts=Xnorm,  
logcounts=Xlog) ) #assays
```

```
reducedDims(SCE) = SimpleList(...)
```

```
SCE@metadata[["clustering"]] = list(...)
```

```
SCE@metadata[["cellType1"]] = vector1
```

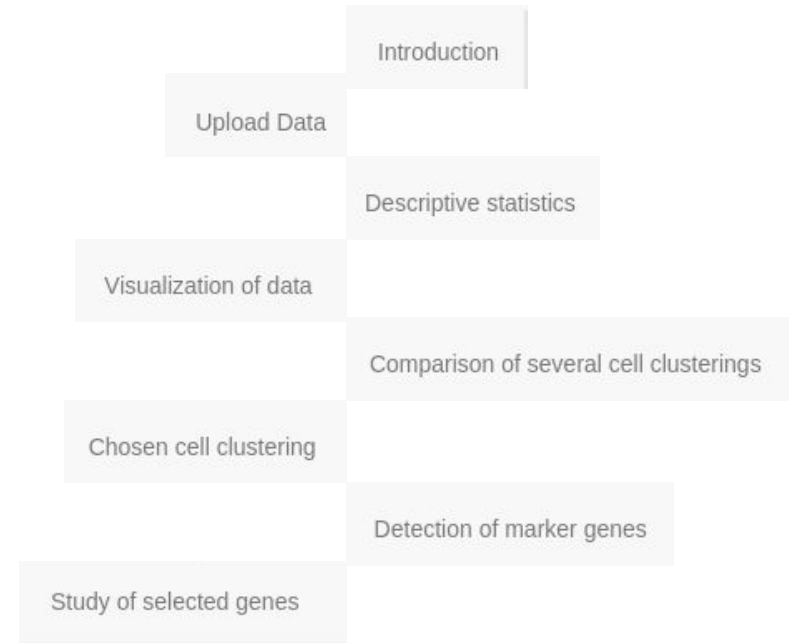
```
SCE@int_elementMetadata$NameType1 <- vectorfeature1
```

# Contents

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## Tabs of the app

- Introduction
- ★ Upload data
- Descriptive statistics
- ★ Visualization of data
- ★ Comparison of cell clusterings
- ★ Analysis of one cell clustering
- ★ Detection of marker genes
- ★ Analysis of genes



# Welcome to this Shiny App

The aim of this shiny application is to exploit clustering results on single-cell RNA-seq data.

This application only uses SingleCellExperiment (SCE) object. The structure of this SingleCellExperiment object, denoted by SCE, is described below.

## Structure of SCE

Please follow these recommendations for building your SCE object :

1. **sce@assays** : contains the different types of data (row counts (counts), normalized counts (normcounts), lognormalized counts (logcounts), ...). By default: 'counts', 'logcounts' and 'normcounts' are required. The rows correspond to features (e.g. genes), the columns to cells.
2. **sce@metadata** contains supplementary information for cells (qualitative variables).
3. **sce@metadata\$clustering** contains a list of the different cell clusterings.
4. **reducedDims(sce)** contains a list of coordinate matrices for each considered dimensionality reduction method.
5. **sce@int\_elementMetadata** contains the supplementary information for features. In particular, sce@int\_elementMetadata\$KnownFunc may contain a binary dataframe where each column correspond to a biological function and some features in row.

This app is compatible with R version  $\geq 3.6.3$  (2020-02-29) and requires the following packages:

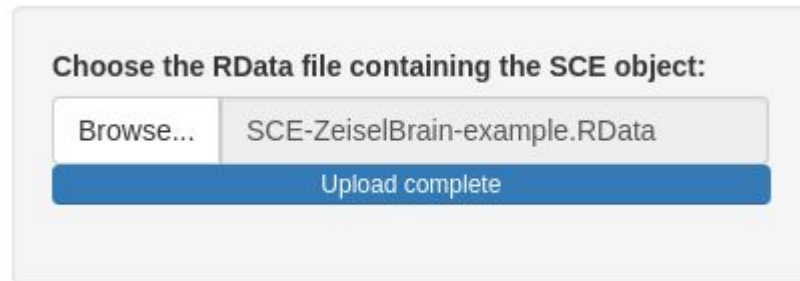
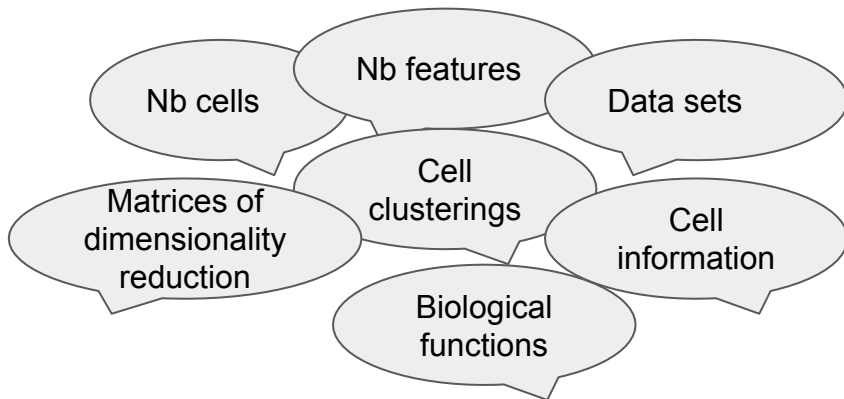
- scales 1.1.0
- Seurat 3.1.5
- dplyr 0.8.5
- scater 1.14.6
- SingleCellExperiment 1.8.0
- DT 0.13
- plotly 4.9.2.1
- ggplot2 3.3.0
- shiny 1.4.0.2

# Upload data

Upload a SCE object in a Rdata file

Does the application detect the information?

User can control if his object is well constructed :



# sc-ASTEC

- Introduction
- Upload Data**
- Descriptive statistics
- Visualization of data
- Comparison of several cell clusterings
- Chosen cell clustering
- Detection of marker genes

Study of selected genes

### Choose the RData file containing the SCE object:

Browse... SCE-ZeiselBrain-example.RData

Upload complete

### The SCE object contains:

- the study of 3005 cells expressed in 2072 features.
- the following data sets:  
counts, normcounts, logcounts
- the following cell clusterings:

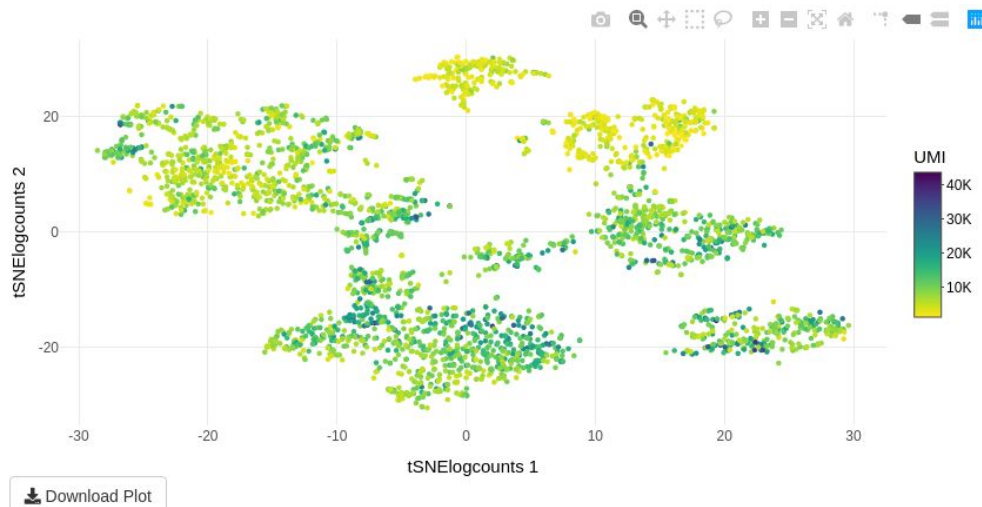
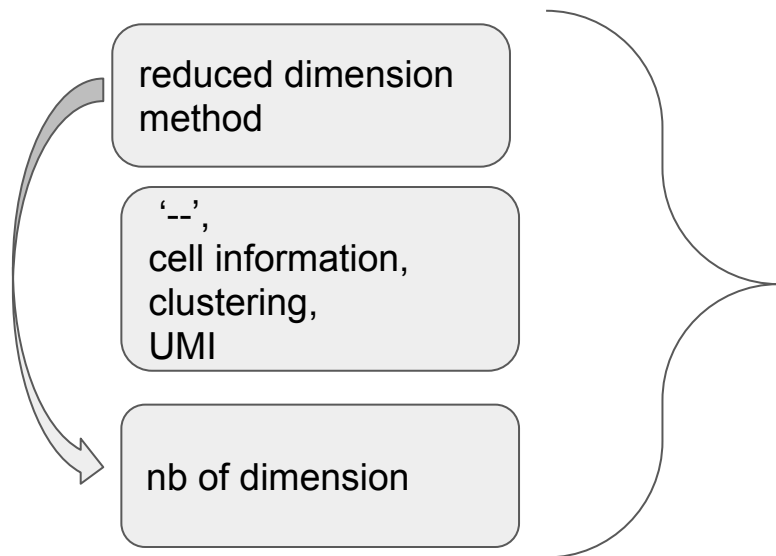
Name	Nb of clusters
ClustZeisel	9
ClustPcared	9
ClustSC3	9
ClustSeurat	15

- the following coordinate matrices of dimensionality reduction methods:

Name	Nb of dimensions
PCAllogcounts	10
tSNElogcounts	2
UMAPlogcounts	2
PCAnormcounts	10

# Visualization in dimensionality reduction

User choices :



# sc-ASTEC

- Introduction
- Upload Data
- Descriptive statistics
- Visualization of data**
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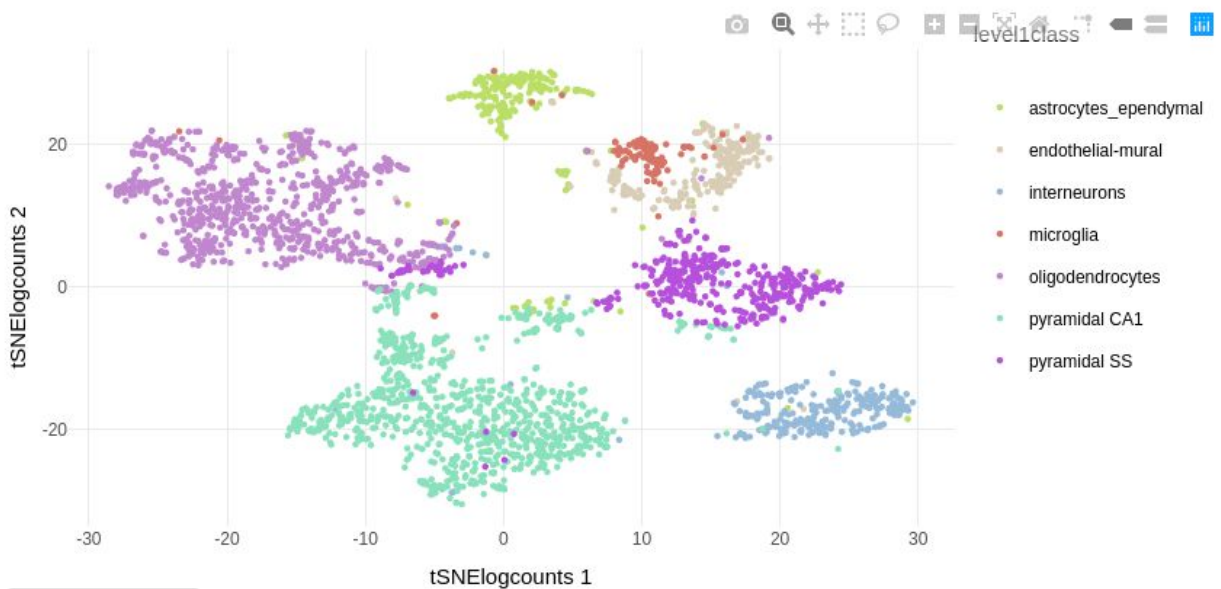
Study of selected genes

**Choose the coordinates of dimension reduction:**  
tSNElogcounts

**Cell information for colour:**  
level1class

**Choose the number of dimensions:**  
2

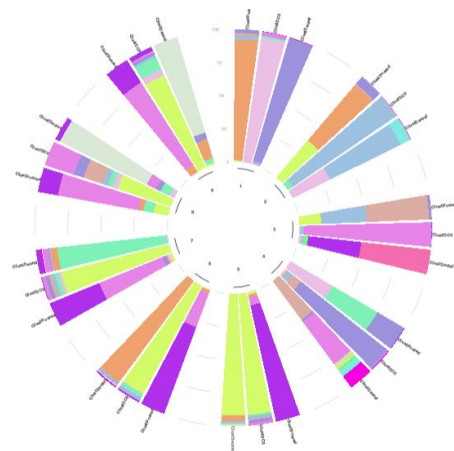
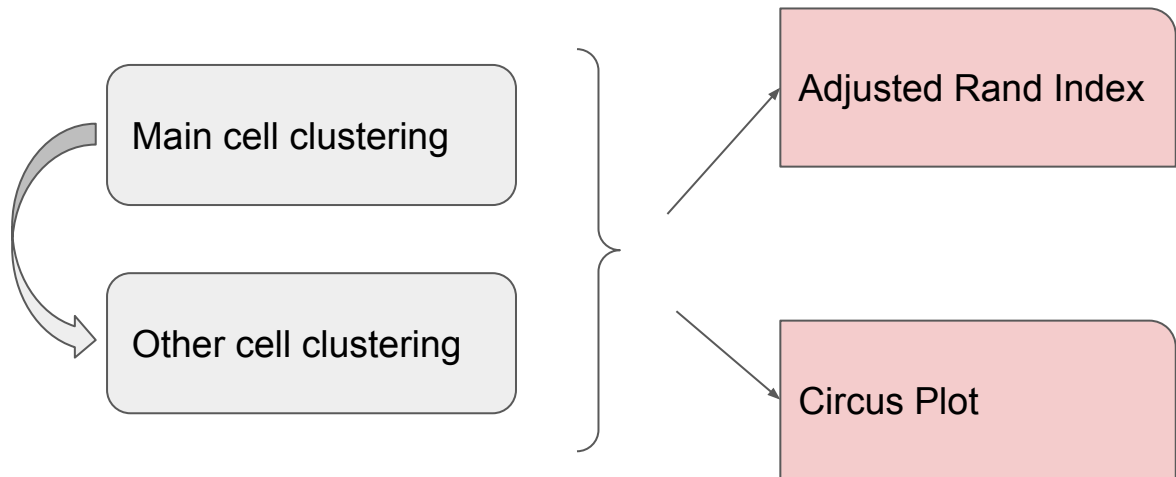
Graph of the cells in a low-dimensional space obtained with the chosen dimensionality reduction method.



[Download Plot](#)



# Comparison of cell clusterings



# sc-ASTEC

- Introduction
- Upload Data
- Descriptive statistics
- Visualization of data
- Comparison of several cell clusterings**
- Chosen cell clustering
- Detection of marker genes

Study of selected genes

Choose the cell clusterings to compare

**The main cell clustering:**

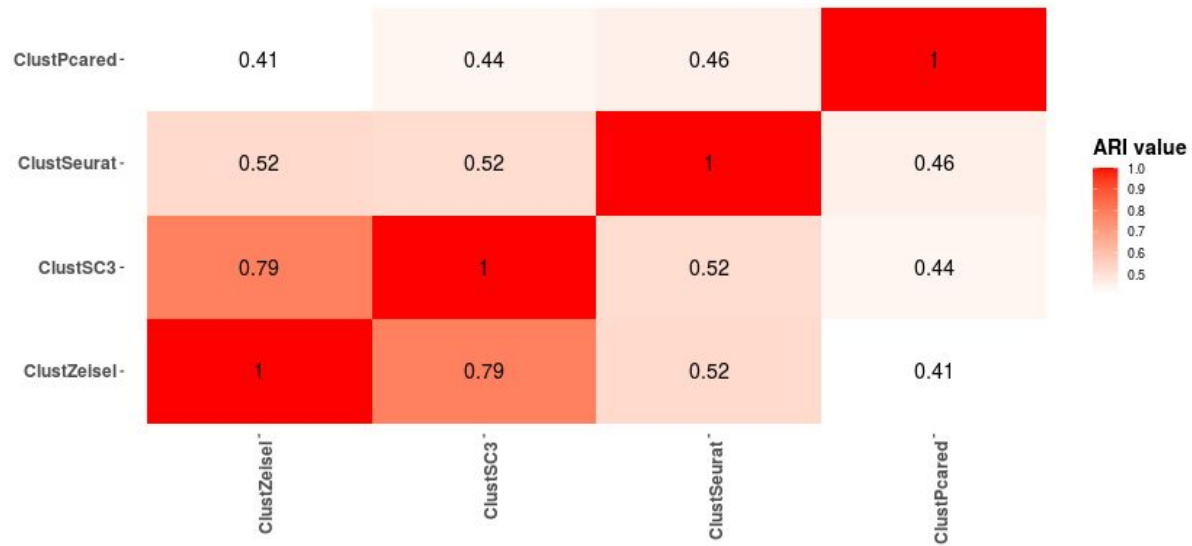
ClustZeisel

**The other cell clusterings:**

ClustSC3 ClustSeurat ClustPcared

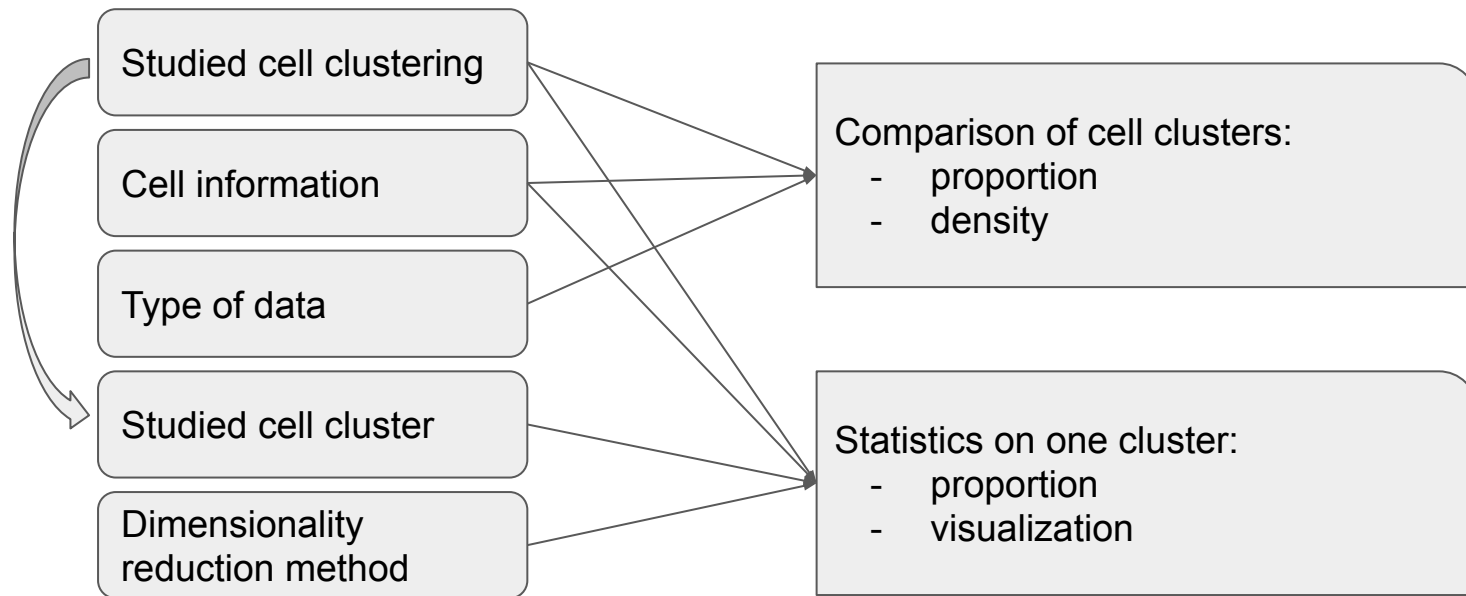
ARI [CircusPlot](#)

Table of adjusted rand index (ARI) between each cell clusterings.



Download Plot

# Analysis of cell clustering



Study of selected genes

Choose the studied cell clustering:

ClustZeisel

Choose cell information:

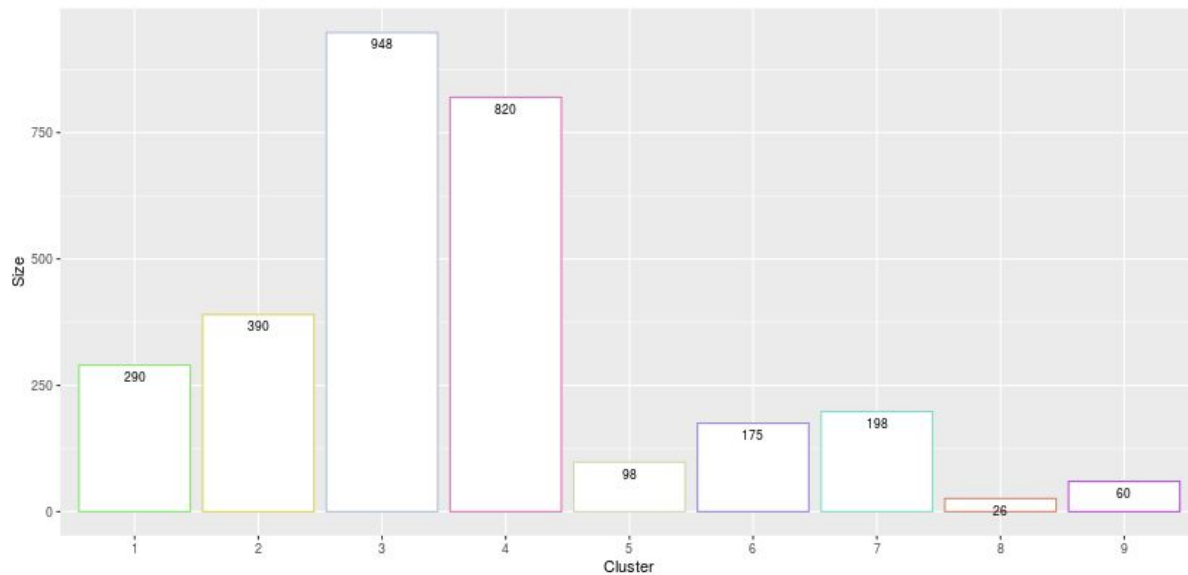
level1class

Choose a type of data:

counts

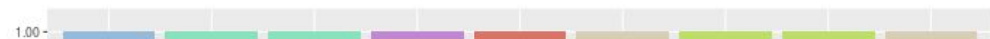
Description | **Specific cluster**

Number of cells per cluster:



Download Plot

Proportion of cell information per cluster:



# Detection of marker genes

Choose two cells groups (based on clusters) [C1, C2]

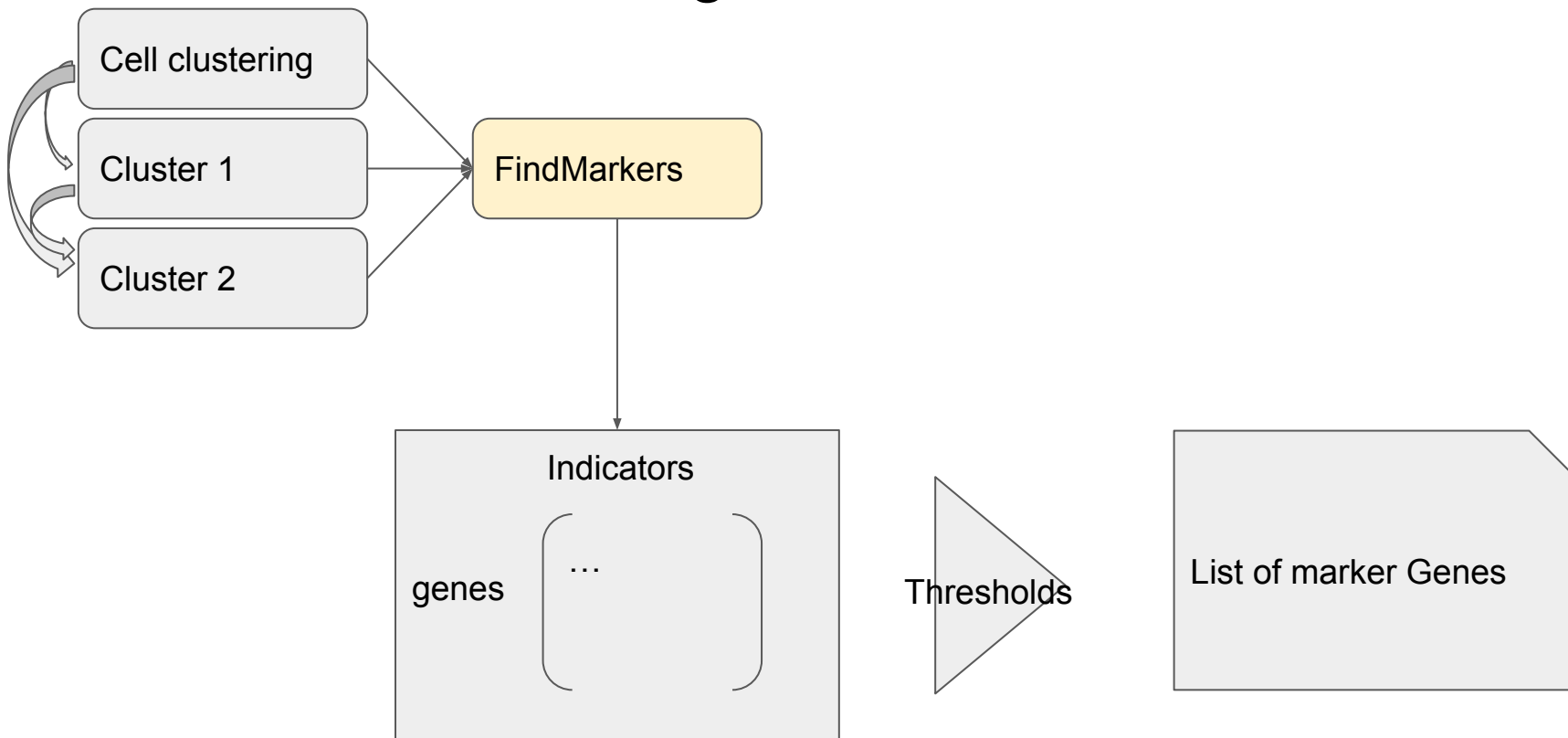
One to one  
One to many  
Many to one  
Many to many

Calculation of indicators by gene:

- **pct.1 (pct.2)**: % of express cells in C1 (resp. C2)
- **avg\_logFC**: log fold-change of the average expression between the two groups
- **AUC**: the area under the ROC curve
- **p\_val\_adj**: the adjusted p-value on *Wilcoxon test*, based on *Bonferroni correction* using all genes in the dataset

Marker genes := Selection with threshold on theses indicators

# Detection of marker genes



Study of selected genes

Choose the studied cell clustering:

ClustZeisel

Choose one (some) studied cluster(s):

6

Choose the comparing group of clusters (one, some or all others):

9

Go - Compute indicators.

Calibration of indicators

Maximum threshold for the adjusted p-val:



Minimum threshold for the logfoldchange:



Find Marker Genes | Indicators

With the chosen thresholds, 10 marker genes are detected.

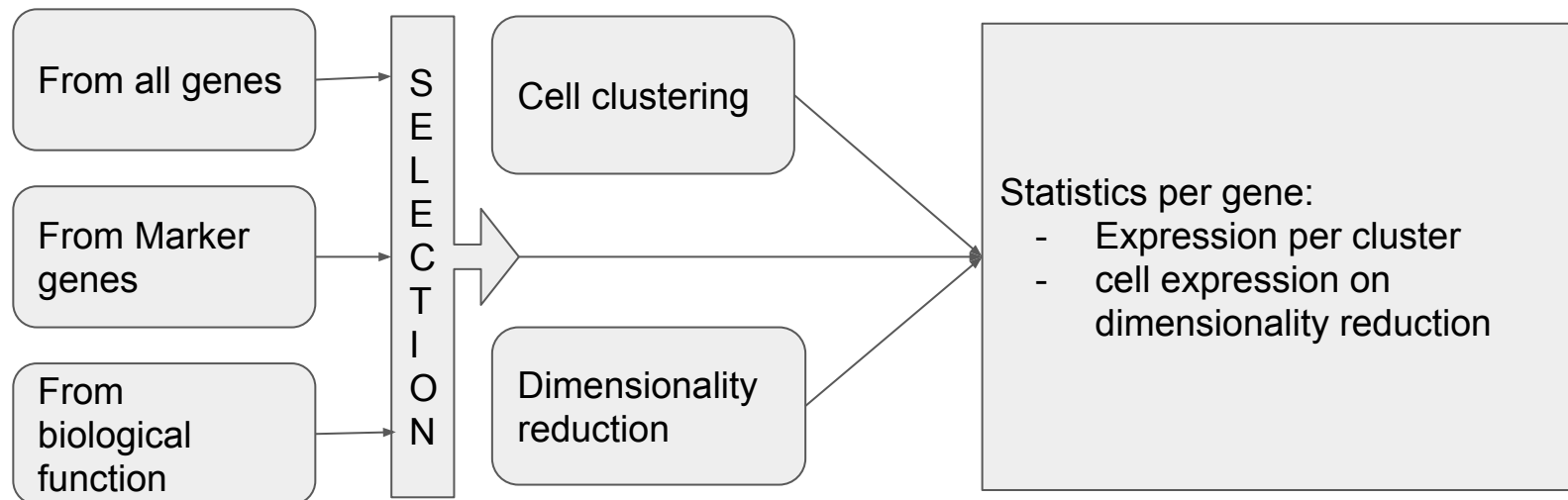
Show 10 entries

Search:

	gene	avg_logFC	pct.1	pct.2	p_val_adj	AUC	delta	func
1	Arap3	2.913	0.754	0.133	0.000	0.833	0.621	
2	Fam101b	2.998	0.560	0.033	0.000	0.767	0.527	
3	Flt1	2.742	0.863	0.133	0.000	0.886	0.730	
4	Gimap6	3.057	0.783	0.100	0.000	0.860	0.683	
5	Itm2a	2.736	0.994	0.333	0.000	0.933	0.661	
6	Ly6c1	2.976	0.914	0.217	0.000	0.893	0.697	
7	mt-Tc	3.026	0.891	0.550	0.000	0.817	0.341	
8	Slc38a3	3.236	0.640	0.033	0.000	0.805	0.607	
9	Slc7a5	2.848	0.749	0.167	0.000	0.821	0.582	
10	Slco1a4	2.755	0.846	0.150	0.000	0.873	0.696	

Showing 1 to 10 of 10 entries

# Study of selected genes





# Study of selected genes

### Choose the source of genes

- From your object
- From detection of marker genes tab
- From a gene function

### Select genes

- All Genes
- Some Genes

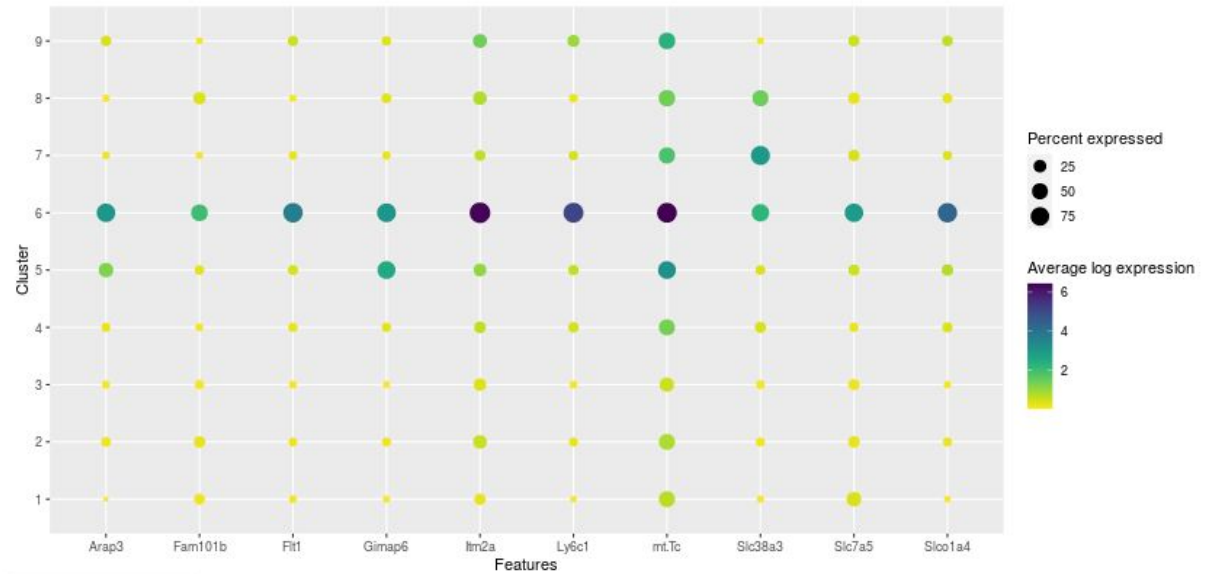
### Choose the studied clustering:

ClustZeisel

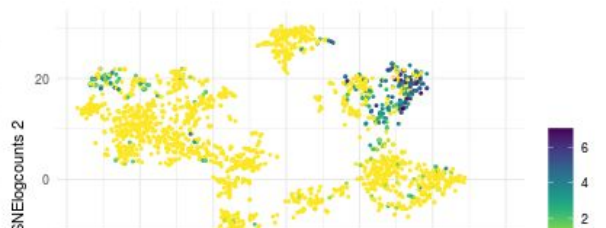
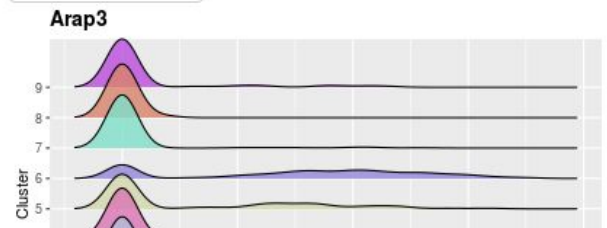
### Choose the dimensionality reduction method:

tSNElogcounts

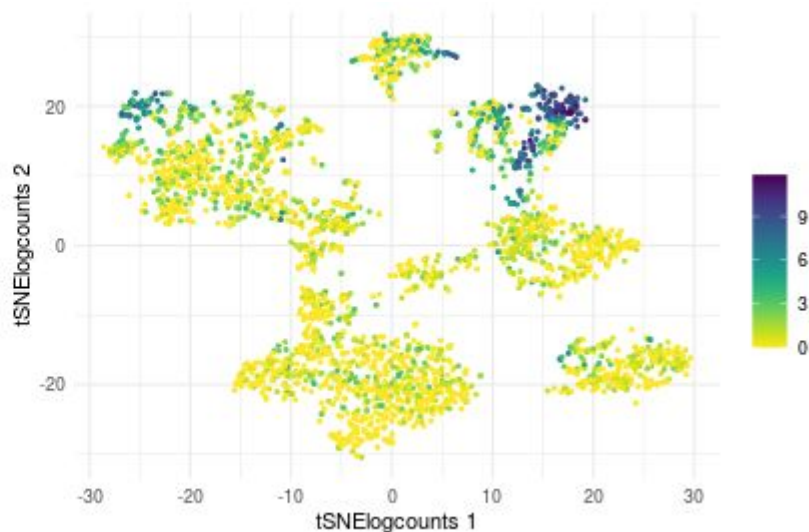
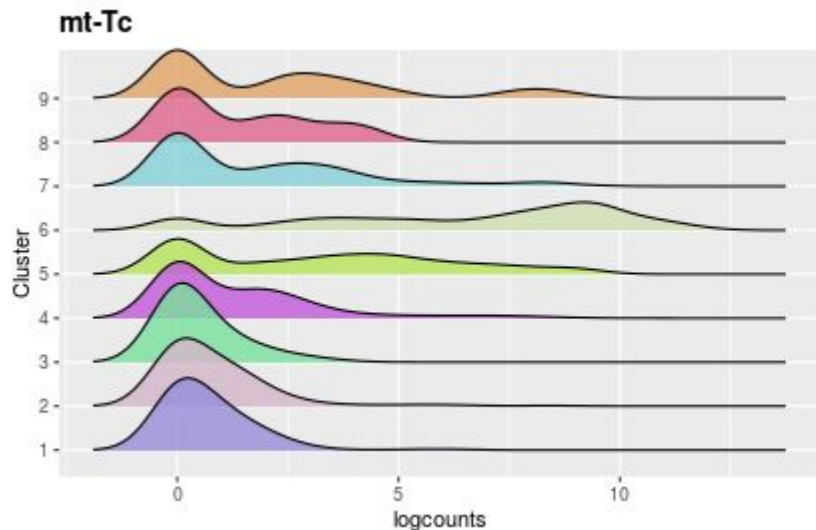
Analyse the selected genes.



Download Plot



# Study of selected genes



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# Advantages

User-friendliness

Many options for analysis

Autonomy

Comparison of cell clusterings

# Improvement

More comparison of cell clusterings

Open structure of sce object

# Deployment

Mathrice : the CNRS server

Access only with internet

The way to use ASTEC-sc without R and packages