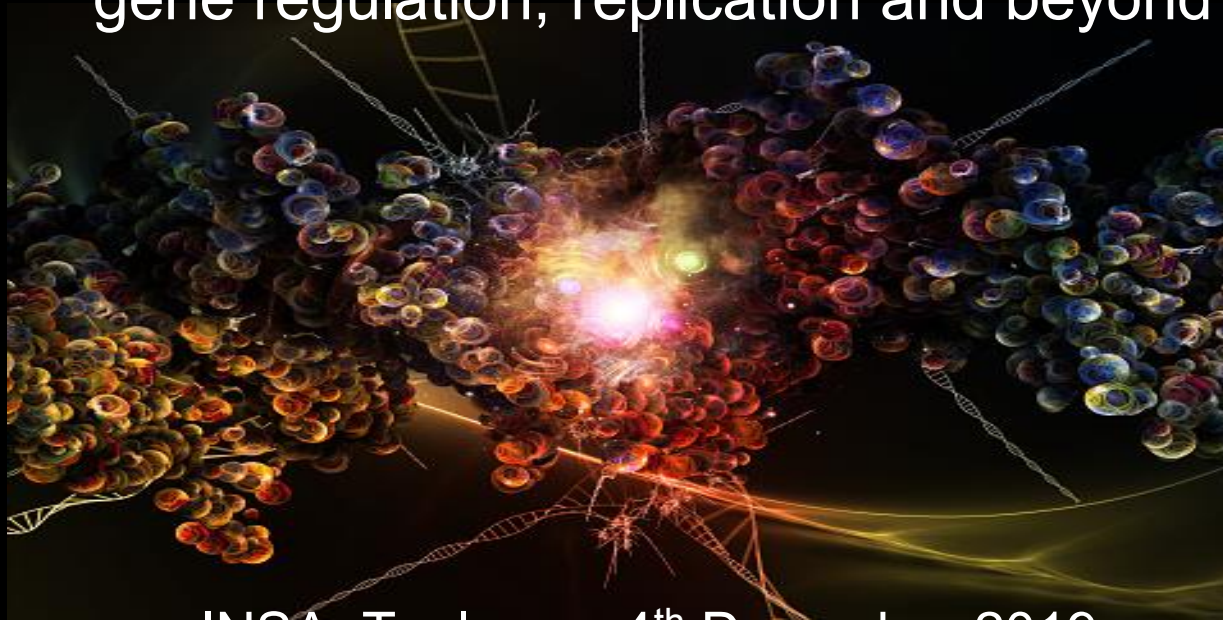


Chromatin 3D organization principles revealed by network theory: gene regulation, replication and beyond



INSA, Toulouse, 4th December 2019

Vera Pancaldi
Cancer Research Center of Toulouse
Barcelona Supercomputing Center



Overview of talk



Networks

Chromatin networks

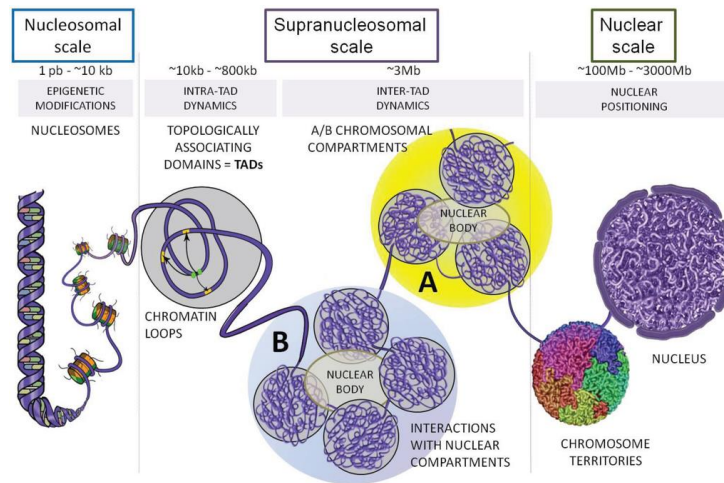
Chromatin Assortativity

Tools

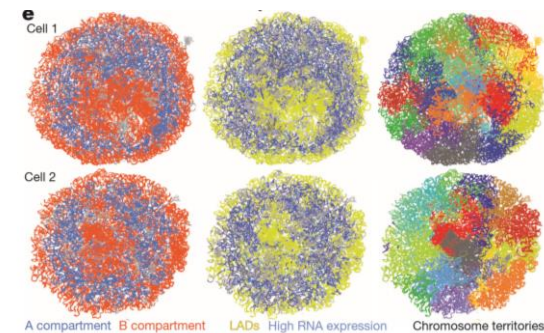
Replication in 3D

Perspectives

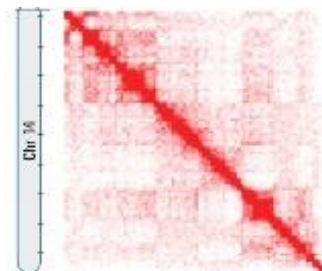
Chromatin 3D structure



Single cell !

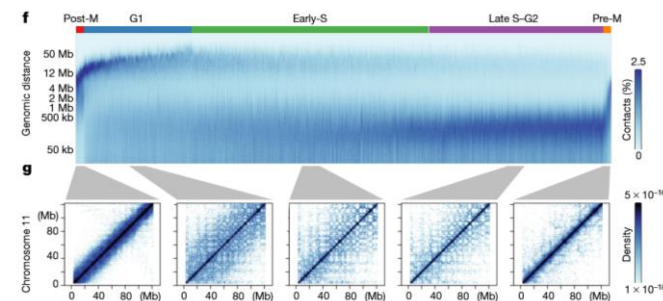


Single cell during the cell cycle



TADs:
More contacts
within
Less outside

Compartments:
Active/Repressed

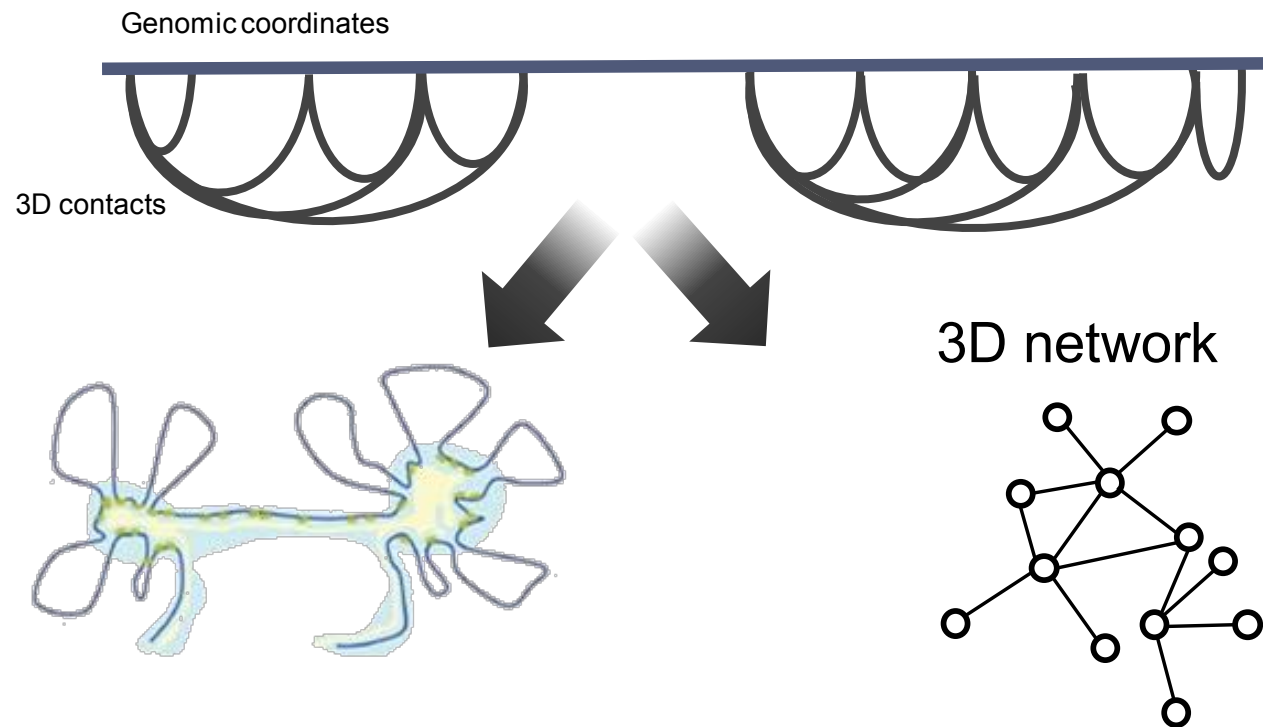


Ea et al. 2015 **Contribution of Topological Domains and Loop Formation to 3D Chromatin Organization**.

Stevens et al. 2017 **3D structures of individual mammalian genomes studied by single-cell Hi-C**

Nagano et al. 2017 **Cell-cycle dynamics of chromosomal organization at single-cell resolution**

Chromatin networks



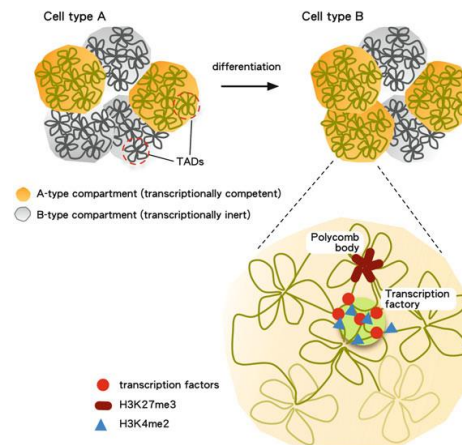
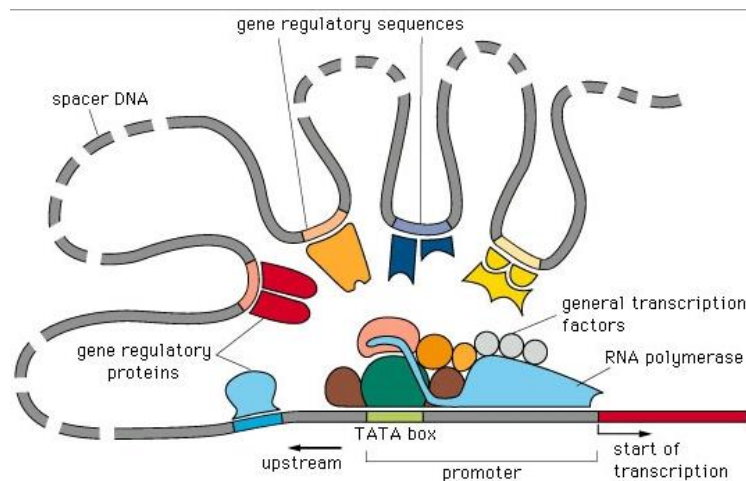
Principal players in gene regulation

Polycomb – gene repression

RNAPII – gene transcription (RNAPIIS2p needed for elongation)

Genes can be co-transcribed (Promoter-Promoter contact, PP)

Regulatory regions bind the gene promoters to activate genes (PO)



What about genes? PCHiC!

Problem so far: Genome wide interaction networks are dominated by interactions far from genes. Need very high coverage to pick promoters and see their interactions.

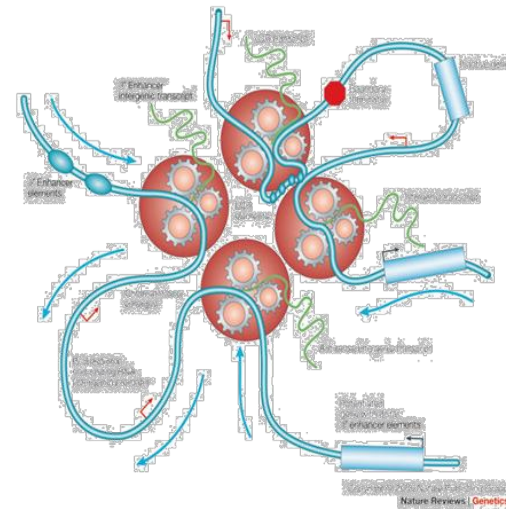
Solution: Promoter-Capture HiC (PCHiC)

Add promoter capture step

To ensure only interactions involving at least one promoter are kept.
(No pull-downs, genome-wide)

Can look for transcription factories:

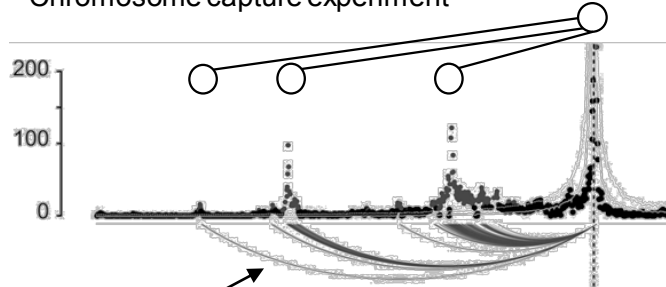
Regions where functionally related transcripts are transcribed



Chakalova et al. 2015, **Replication and transcription: Shaping the landscape of the genome**; Schoenfelder, S. et al. 2015, **The pluripotent regulatory circuitry connecting promoters to their long-range interacting elements**.

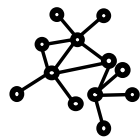
The 3D genome as a network

Chromosome capture experiment

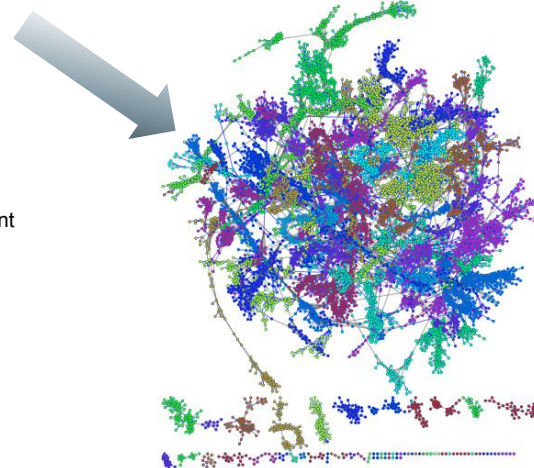


Genomic contacts

3D network



● Genomic fragment



Some other network approaches to chromatin:

Botta et al. 2010, **Intra- and inter-chromosomal interactions correlate with CTCF binding genome wide**

KS Sandhu et al. 2012, **Large-scale functional organization of long-range chromatin interaction networks**

Boulos et al. 2017 **Multi-scale structural community organisation of the human genome**

Mourad et al. 2017 **Uncovering direct and indirect molecular determinants of chromatin loops using a computational integrative approach**

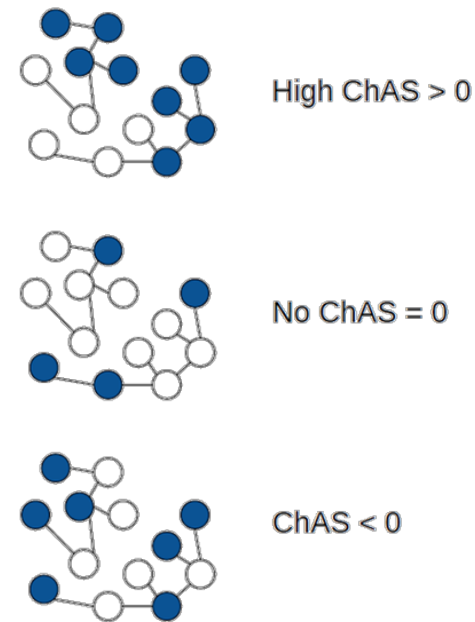
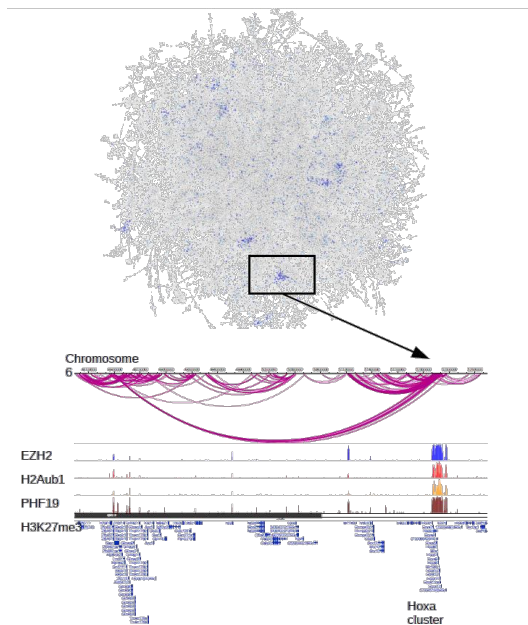
Norton et al. 2018 **Detecting hierarchical genome folding with network modularity**

Pancaldi et al. 2016 **Integrating epigenomic data and 3D genomic structure with a new measure of chromatin assortativity.**

Chromatin Assortativity of epigenetic marks

Project ChIP-seq datasets on 3D chromatin interaction network
PCHiC networks in mouse Embryonic Stems Cells mESCs)
(Collaboration with Peter Fraser, Babraham Institute)

Do regions with specific marks cluster?
Inspired by social networks (the twitter story)
Define Chromatin Assortativity (ChAs)



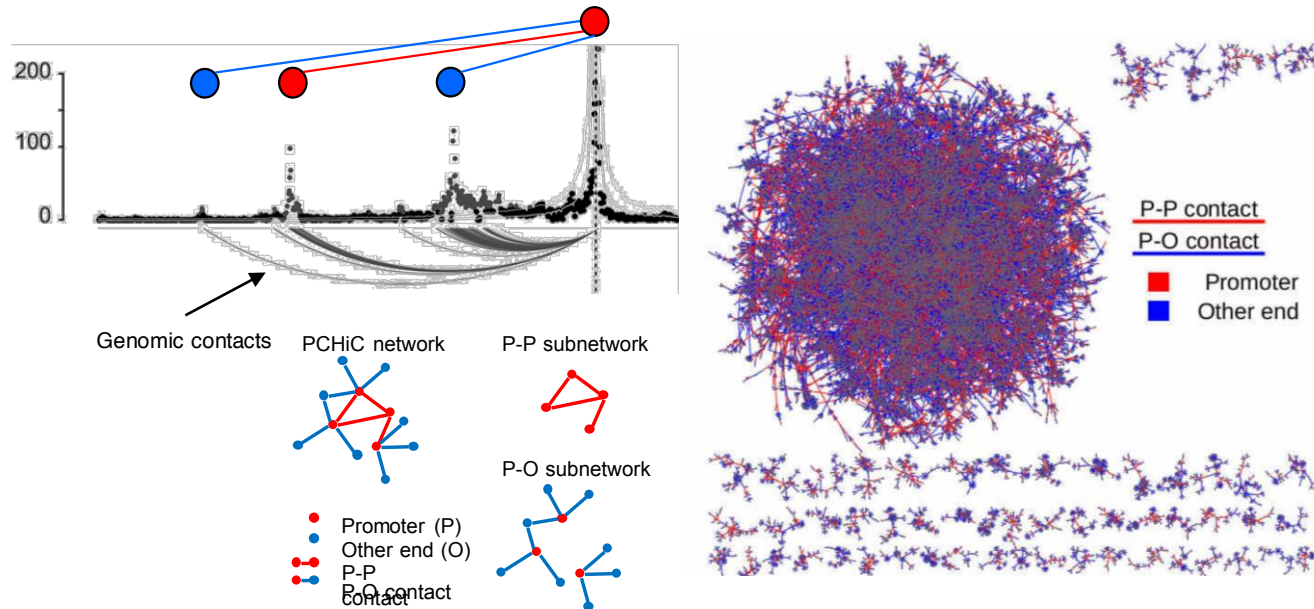
Pancaldi et al. 2016 Integrating epigenomic data and 3D genomic structure with a new measure of chromatin assortativity.

Promoter Capture HiC networks in mESC

Nodes are chromatin fragments (5kb median size)

Connections (edges) are 3D contacts

Significant contacts are detected using CHiCAGO



Schoenfelder et al. 2015 **The pluripotent regulatory circuitry connecting promoters to their long-range interacting elements.**

Cairns et al. 2016 **CHiCAGO: robust detection of DNA looping interactions in Capture Hi-C data.**

Comparing ChAs in P-P and P-O subnetworks

Identify features that have different ChAs in P-P and P-O contacts in mESC

PCG - on diagonal

Similar ChAs >0 in P-P and P-O

Equal importance

RNAPII - spread

Variable ChAs in P-O,

ChAs >0 in P-P

H3K4me3

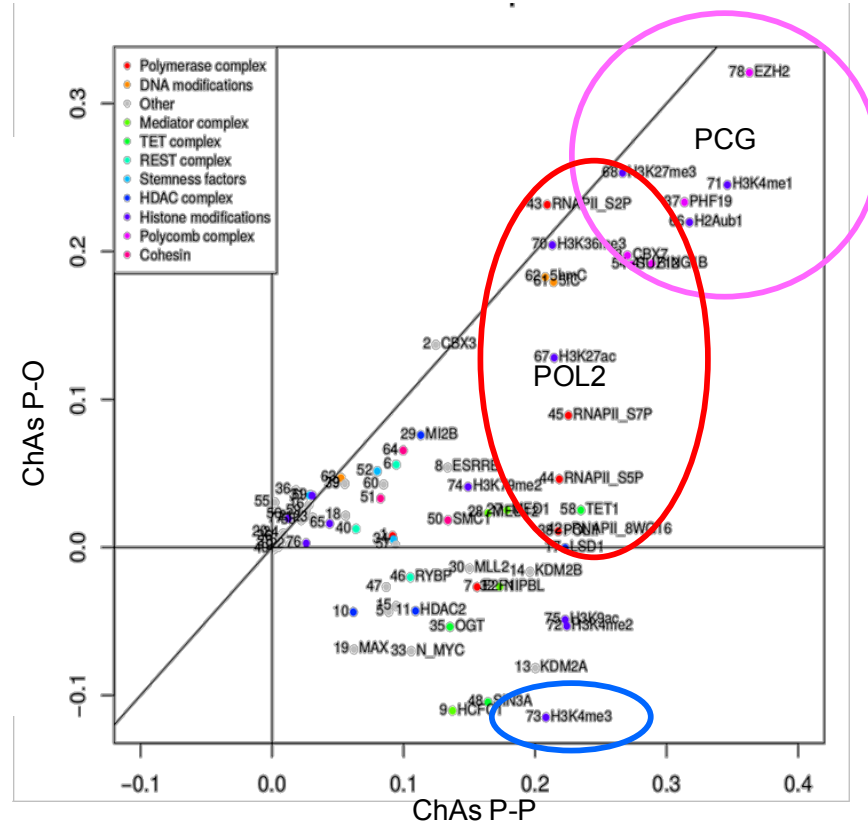
ChAs >0 in P-P

ChAs <0 in P-O

(only present in promoters)

Fragments that have this mark are more likely to interact

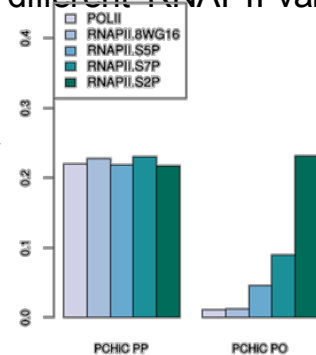
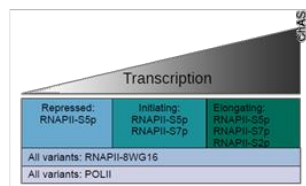
→ **Preferential contacts of active gene promoters**



Assortativity of RNAPII

5 Different RNAPII features

Binding peaks for different RNAPII variants

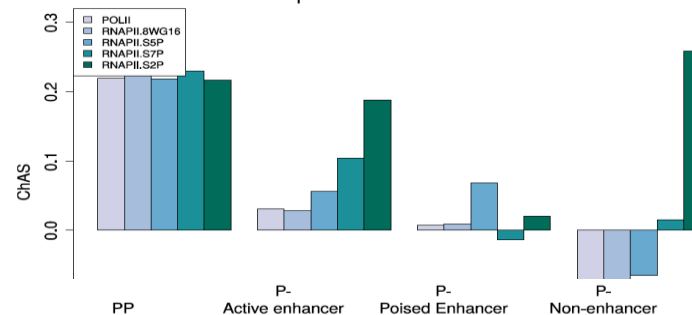


ChAs of RNAPII in P-O variable

Non-elongating RNAPII has low ChAs in P-0

Is expression enhancing mediated by RNAPII S2p?

Assortativity of RNAPII variants in Interactions of promoter and enhancers



Active enhancer
H3K4me1+H3K27ac

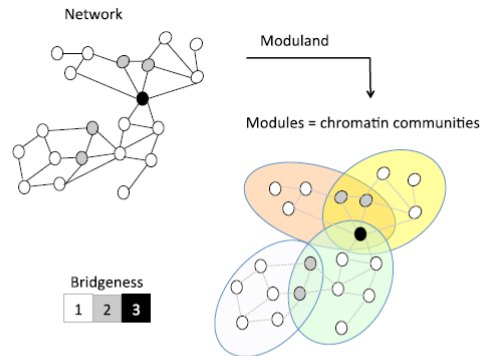
Poised Enhancer
H3K4me1+
H3K27me3

Non-enhancer
No H3k4me1

Chromatin network analysis

Apply Moduland (Cytoscape plugin) to identify overlapping chromatin communities, measure bridgeness

	Bridgeness	Betweenness centrality	Degree	CC	Party/date
PCG	Low	High	High	Very low	PARTY
RNAPII general	High	Low	Low	Low	DATE
RNAPII S2p	Low	Very low	Very low	Medium	NOT HUB



Interpretation:

PCG is a stable hub (across cells, in time?)

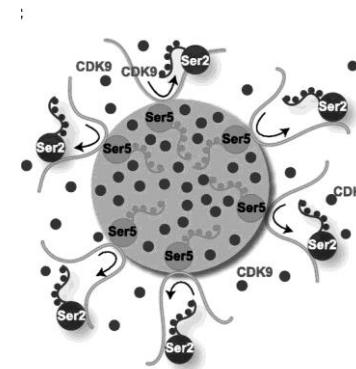
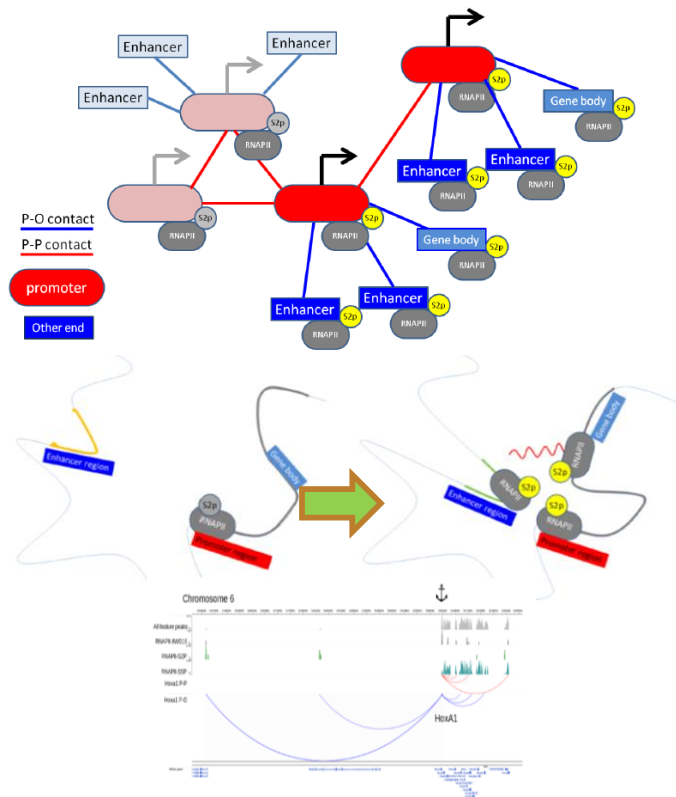
RNAPII general dynamic (reflecting transcription regulation?)

RNAPII S2p peripheral

LCC= Large connected component

The model

Whereas RNAPII S5P accumulates in transcription factories, RNAPII S2p stays peripheral



A model of transcription; gene promoters are loaded with RNAPII-Ser5P (Ser5 light gray) in factories. Elongating RNAPII S2p (Ser2, dark gray) moves to the adjacent nuclear space when it becomes phosphorylated at Ser2 by CDK9

A. Ghamari et al. In vivo live imaging of RNA polymerase II transcription factories in primary cells *Genes Dev.* 2013;27:767-777

Ghavi-Helm et al. Enhancer loops appear stable during development and are associated with paused polymerase. *Nature.* 2014;512:96-100.

Pancaldi et al. 2016 Integrating epigenomic data and 3D genomic structure with a new measure of chromatin assortativity.

Other applications

GARDEN-NET

Genome ARchitecture Data, Epigenome and Nucleome - Network Exploration Tool

<https://pancaldi.bsc.es/garden-net>



Here you will be able to visualize chromosome conformation capture datasets as networks of interacting chromatin fragments. The published datasets available were generated with the *Promoter Capture HiC* technique, which returns contacts involving promoters. For more information see: [GARDEN-NET and ChAseR: a suite of tools for the analysis of chromatin networks](#)

Left click on nodes to navigate to WashU browser.
Right click on nodes to zoom into their neighborhood.



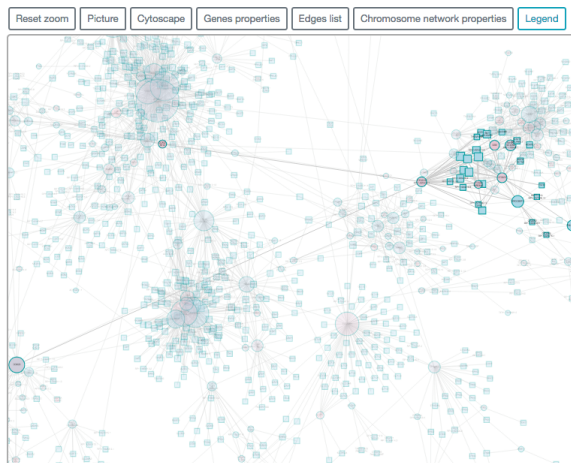
Homo sapiens : Neutrophils

Submit

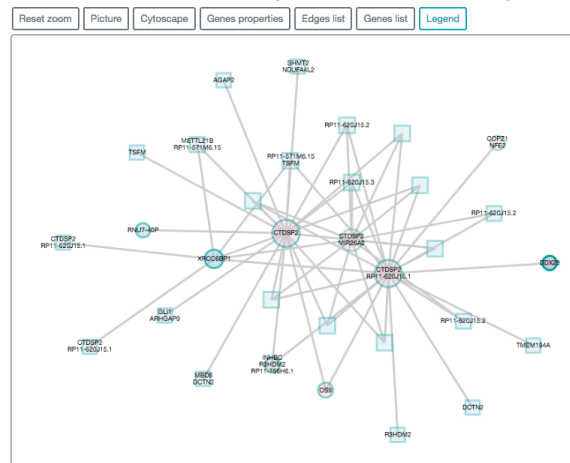
GARDEN-NET uses functionality provided by the *ChAseR* R package to integrate datasets and compute chromatin assortativity (*Pancaldi et al. 2016*). User submitted features can also be visualized on the networks and used for the network based calculations.

A selection of epigenomic features that have been mapped to the chromatin fragments will be available from the drop-down menu on the right. Select one of them to visualize chromatin fragments that have that feature and to calculate statistics relating to this feature and the 3D network.

Chromosome 12



CTDSP2 MIR26A2 (12:58212900-58223142)



Data sources

Chromosomes

12 -

Features

GeneExp -

GeneExp statistics	Values
Abundance	0.370
ChAs	-0.230
Random ChAs interval	-0.19,-0.161
Mean degree	12.370
Abundance (PP)	2.450
ChAs (PP)	0.202
Mean degree (PP)	3.630
Abundance (PO)	0.340
ChAs (PO)	-0.326
Mean degree (PO)	10.680

Upload features

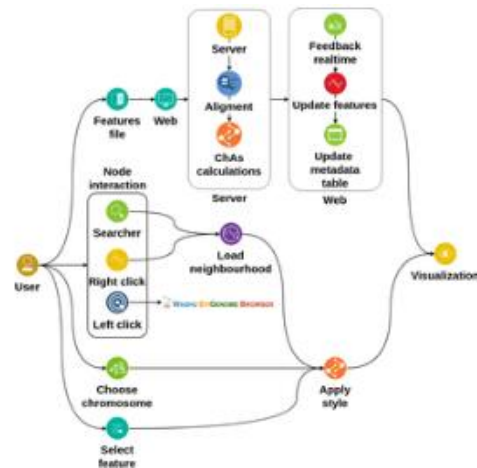
Madrid*, Raineri* and Pancaldi, 2019 GARDEN-NET and ChAseR: a suite of tools for the analysis of chromatin networks
BioRxiv 717298, (submitted)

GARDEN-NET

Genome ARchitecture Data, Epigenome and Nucleome - Network Exploration Tool



Interactive and processing in real time



Technologies

Web:

- Yarn (package management)
- Webpack (bundler manager)
- React (Router, Reactrap) with Redux
- Cytoscape.js (network visualization)
- Bootstrap (style)



Server:

- Nginx (HTTP server)
- Python with Flask (REST API)
- R with (ChAseR, Igraph, GenomicRanges, rjson, Tidyverse)
- Bash with pipes, sed & gnu parallel (pipeline integration)
- Celery with redis (task queue system)
- Docker & Docker Compose



Chromatin Contact networks

Promoter Capture HiC data for haematopoietic cells in human. [Javierre et al. 2016](#)

Promoter Capture HiC data for mouse embryonic stem cells [Schoenfelder et al. 2015](#)

Features

Mouse embryonic stem cells histone modifications and 78 ChIP-Seq datasets. [Juan et al. 2016](#)

GeneExp from [Finotello et al. 2019](#)

GeneExpEPIVAR for Monocytes, Neutrophils and Tcells from [Chen et al. 2016](#)

Human Histone modification data: EPIVAR from [Chen et al. 2016](#)

Human Replication Timing data (GM12878). [Pope et al. 2014](#)

PCHiC data processed with CHICAGO. [Cairns et al. 2016](#)

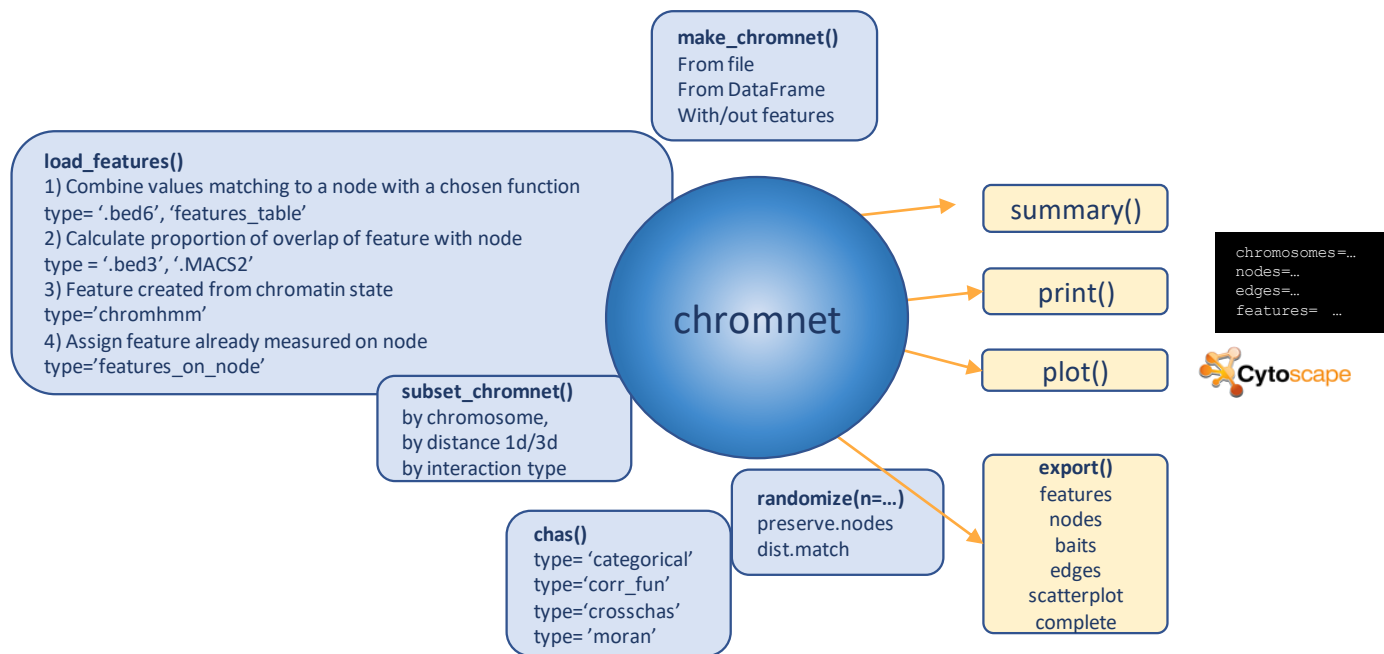
Technical details at <https://github.com/VeraPancaldiLab/GARDEN-NET>

Madrid*, Raineri* and Pancaldi, 2019 GARDEN-NET and ChAseR: a suite of tools for the analysis of chromatin networks
BioRxiv 717298, (submitted)

ChAseR: an R package



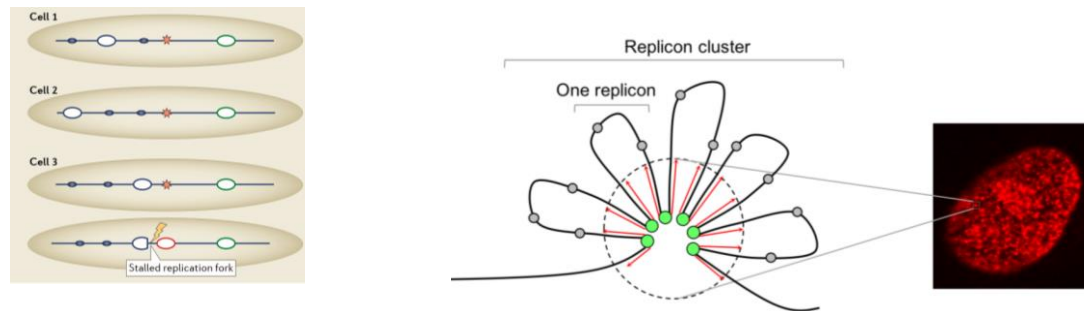
<https://bitbucket.org/eraineri/chaser/>



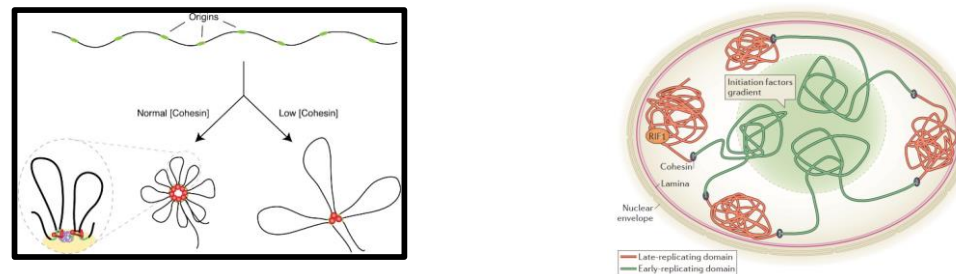
Madrid*, Raineri* and Pancaldi, 2019 GARDEN-NET and ChAseR: a suite of tools for the analysis of chromatin networks
BioRxiv 717298, (in revision)

Mammalian DNA replication in 3D

Stochastic firing model: constitutive, flexible, dormant origins



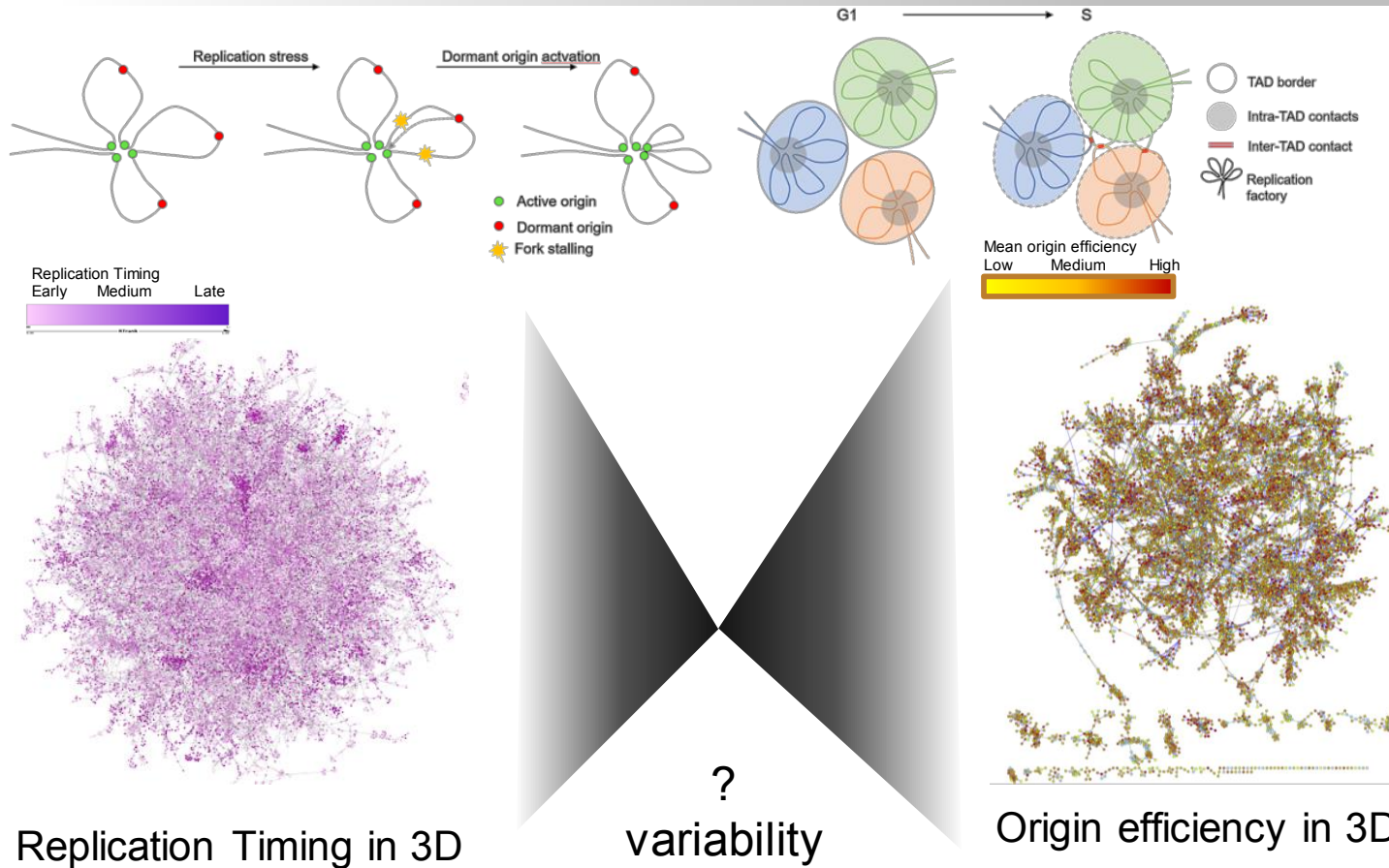
Cohesin might mediate replicon loops, which assemble in Replication Domains (coinciding with TADs)



Fragkos et al. **DNA replication origin activation in space and time**. Nat. Rev. Mol. Cell Biol. 2015

Guillou, E. et al. **Cohesin organizes chromatin loops at DNA replication factories**. Genes Dev. 2010

A global view of replication in 3D in mouse

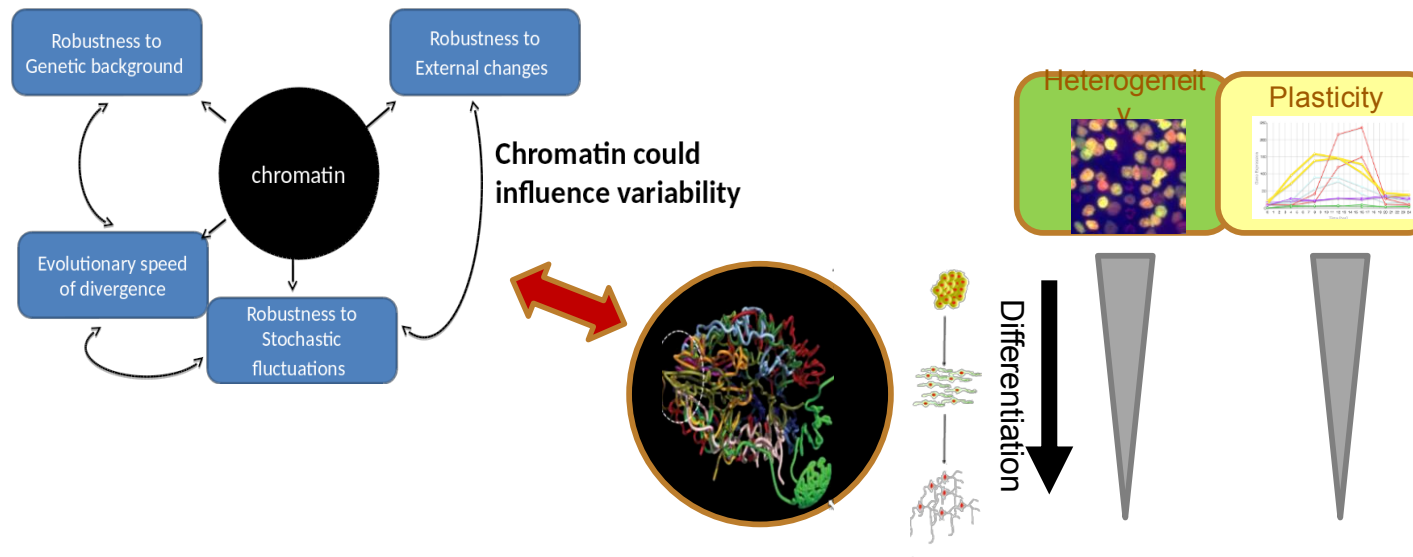


Perspectives

Chromatin, heterogeneity, plasticity, stemness...

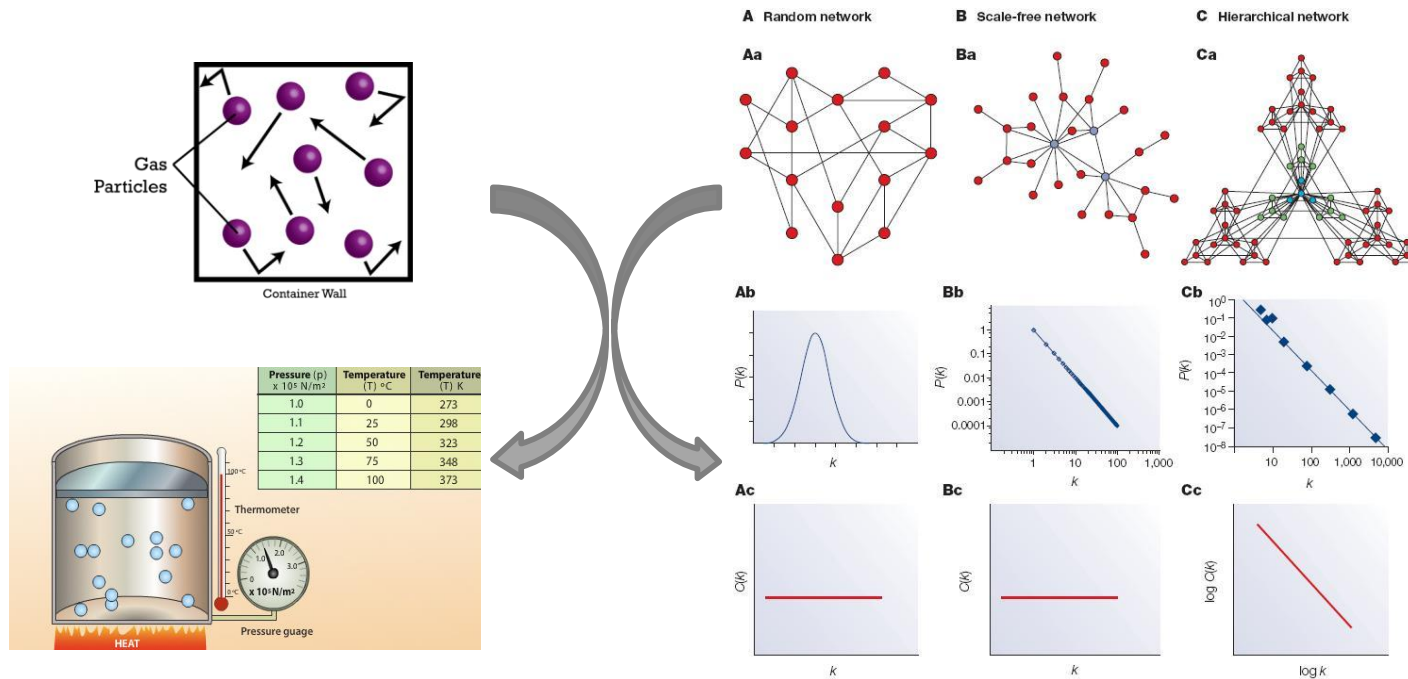
Chromatin state (methylation/histone modifications etc...) can affect

- Plasticity (rapidly regulated stress genes)
- Single cell heterogeneity (noisy promoters)
- Inter-individual differences
- Evolutionary divergence speed



A systems approach

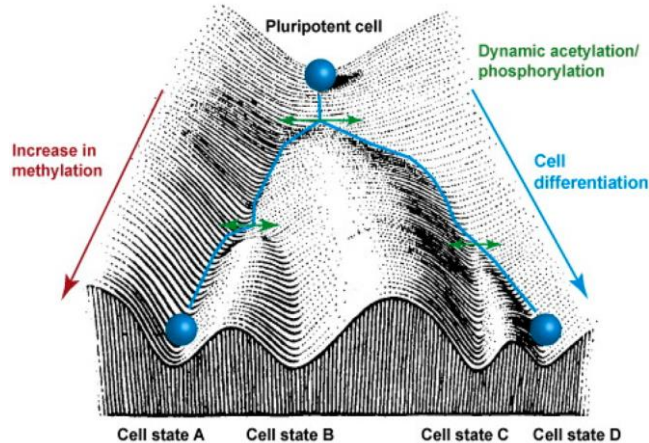
Thermodynamics: from particles' positions and velocities to pressure and temperature



Network theory: from nodes and edges to degree distribution, clustering coefficient, ...

Interdisciplinary approach: borrow concepts from studies on other networks

Cellular differentiation and response



Isogenic heterogeneity

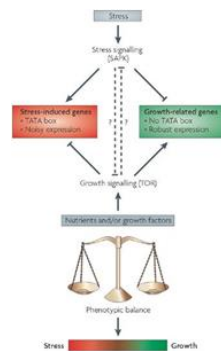
Large fluctuations, motion across landscape

Plasticity – in response to external signals

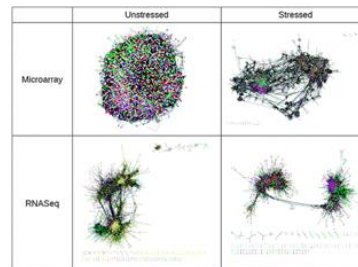


Reduced heterogeneity/plasticity

Well-defined phenotype



Expression Correlation network



System level Stress Response
Network disaggregates into modules
Increase in heterogeneity

Tuning gene expression to changing environments: from rapid responses to evolutionary adaptation López-Maury L et al. *Nat. Rev. Gen.* 2008.

Pancaldi V, Schubert F and Bahler J. **Meta-analysis of genome regulation and expression variability across hundreds of environmental and genetic perturbations in fission yeast.** *Mol. BioSyst.*, 2010.

Stress induces remodelling of yeast interaction and co-expression networks Lehtinen S, Marsellach FX, Codlin S, Schmidt A, Clément-Ziza M, Beyer A, Bähler J, Orengo C, Pancaldi V. *Mol. BioSyst.*, 2013.

MERCI!



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Enrique Carrillo

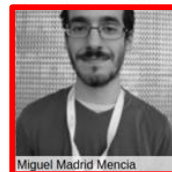
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Pierre Fabre

